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ATP formation coupled to photosynthetic NADP⁺ reduction with artificial electron donors

The photosynthetic reduction of NADP+ by isolated chloroplasts is coupled stoichiometrically to ATP formation¹. The rate of NADPH formation is therefore increased when the phosphorylating system (ADP-P₁-Mg²+) or an uncoupler (NH₄Cl) is added². When NADP+ is reduced at the expense of an artificial electron-donor system such as DCIP-ascorbate³ or diaminodurol-ascorbate⁴ while the oxygen-evolution system is blocked by a specific inhibitor like DCMU, ATP is also formed⁴-6. However, it has been questioned⁻¹,² whether the ATP is formed during the electron transport from the electron donor to NADP+ or rather by a superimposed cyclic photophosphorylation.

We wish to report experiments which strongly support the view that NADP+ reduction at the expense of diaminodurol-ascorbate is indeed directly coupled to ATP formation. This is based on the findings that: (1) the rate of NADPH formation in the diaminodurol system (and to a lesser extent in the DCIP system as shown by Keister¹⁴) is increased by the addition of the phosphorylating system or by NH₄Cl, and (2) the coupled NADP+ reduction is inhibited by phlorizin, a recently described inhibitor⁸ of photophosphorylation. This inhibition by phlorizin is annulled by NH₄Cl.

As Table I shows, NADP+ reduction at the expense of diaminodurol or DCIP,

TABLE I stimulation of NADP+ reduction in three electron-donor systems at pH $_{7.4}$ by NH $_4$ Cl and phosphorylating system and inhibition by phlorizin

The reaction mixture contained in 3 ml: 80 μ moles Tris buffer, 6 μ moles NADP⁺, 10 μ moles ascorbate, broken chloroplasts (P_{1s1}) from spinach with 0.2 mg chlorophyll and excess ferredoxin. 10 μ moles ADP and inorganic phosphate and 5 μ moles MgCl₂ were added where indicated. Illumination with 35000 lux at 15° in N₂.

Additions to 2·10 ⁻⁵ M DCMU	Electron donor (0.2 μmole)	Diaminodurol 10 min		DCIP 15 min		TMPD 15 min	
	Time of illumination						
		NADPH	ATP	NADPH ATP		NADPH ATP	
		(µmoles formed)					
		2.6		1.1		1.5	
+ 10 ⁻³ M NH ₄ Cl		3.6		3.1		1.7	
$+ ADP-P_1-Mg^{2+}$		3.I	3.0	1.5	0.7	1.9	<0.1
$\begin{array}{l} + \mathrm{ADP-P_{i-M}g^{2+}} \\ + \mathrm{10^{-3}Mphlorizin} \\ + \mathrm{ADP-P_{i-M}g^{2+}} \end{array}$		2.6	1.1	0.9	<0.1	2,2	<0.1
+ 10 ⁻³ M phlorizin + 10 ⁻³ M NH ₄ Cl		3.3	0.6	2.4	0.2	2.2	<0.1

Abbreviations: DCIP, 2,6-dichlorophenolindophenol; DCMU, 3(3,4-dichlorophenyl)-1,1-dimethylurea; TMPD, N,N,N',N'-tetramethyl-p-phenylenediamine.

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kept reduced by ascorbate, is stimulated by the addition of the uncoupler NH4Cl or by the phosphorylating system (ADP-P_i-Mg²⁺). The coupled NADP+ reduction is inhibited by 10⁻³ M phlorizin down to the rate of the basal (non-phosphorylating) electron transport (as Izawa, Winget and Good have described for the ferricyanide system). This inhibition by phlorizin is annulled by the uncoupler NH₄Cl. In the TMPD system, however, which at this concentration of TMPD is not coupled to ATP formation^{8,10}, NADP+ reduction is only slightly stimulated by NH₄Cl and is not inhibited by phlorizin. Basal as well as coupled electron transport in the diaminodurol system is higher than in the DCIP, and markedly higher than in the TMPD system, showing again the superiority of diaminodurol as electron donor⁴. The high basal rate in the diaminodurol system is probably due to a bypassing of the rate-limiting phosphorylation site, as diaminodurol has indeed two points of entry into the electrontransport chain4. In the presence of the phosphorylating system diaminodurol uses the point of entry before the phosphorylation site, since ATP is now formed stoichiometrically with respect to NADPH and the rate of the electron transport has been increased.

The experiments in Table I were run at pH 7.4 since at this pH the stimulation of the electron transport by an uncoupler or the phosphorylating system is more pronounced². As Table II shows, the stimulation of coupled NADP⁺ reduction with water as the electron donor (and therefore with oxygen evolution) is markedly increased by the addition of NH₄Cl at pH 7.4 but very little at pH 8.0. The stimulation of basal (non-phosphorylating) NADP⁺ reduction with an artificial donor system (diaminodurol–ascorbate) by NH₄Cl or phosphorylating system is also more clearly seen at pH 7.4 than at pH 8.0 (Table II).

TABLE II STIMULATION OF NADP+ REDUCTION BY NH₄Cl and phosphorylating system at different pH Experimental conditions as in Table I; 10 min light.

Additions	pH 7.4		рН 8.o		
	μmoles NADPH formed	μatoms oxygen evolved	μmoles NADPH formed	μatoms oxygen evolved	
Water as electron donor					
$+ ADP-P_1-Mg^{2+}$	1.7	1.4	4.2	3.8	
+ ADP-P ₁ -Mg ²⁺ + 10 ⁻³ M NH ₄ Cl	4.4	3.8	4.6	4.5	
+ 2·10 ⁻⁵ M DCMU + 0.2 µmole diaminodur	rol				
· · · · ·	2.2		3.0		
+ 10 ⁻³ M NH ₄ Cl	3.1		3.1		
$+ ADP-P_i-Mg^{2+}$	3.1		3.3		
$+ ADP-P_1-Mg^{2+} + 10^{-3} M NH_4Cl$	3.6		3.6		

The rate of NADP+reduction at pH 7.4 (but not at pH 8.0) in the presence of phosphorylating system is even further increased by NH_4Cl when O_2 is evolved but not when diaminodurol is the electron donor (Table II). This shows that coupled electron transport at pH 7.4 with water as the electron donor is limited by the ATP-formation system which has a pH optimum at pH 8.0; the electron transport is

therefore markedly stimulated by the further addition of an uncoupler (Table II) (as is well known for the ferricyanide system^{11–13}). The DCIP system is also more stimulated by an uncoupler than by the phosphorylating system at pH 7.4 indicating that the ATP-forming system is limiting the electron transport (Table I). In the diaminodurol system, however, NH₄Cl and the phosphorylating system give the same stimulation of the basal electron transport (Tables I and II) and further addition of NH₄Cl to the phosphorylating system has only a slight effect (Table II), indicating that the ATP-forming system is not or less limiting. NADPH formation with water or with diaminodurol as the electron donor is coupled to a stoichiometric ATP formation. The absolute rate of ATP formation during NADP+ reduction at pH 7.4 with diaminodurol as the donor is therefore higher than with water as the electron donor. This might be taken as an indication that in the diaminodurol system at pH 7.4 another phosphorylating site is participating which has an optimum at a lower pH.

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On the possible role of structural protein in the binding and translocation of adenine nucleotides in mitochondria

The permeability studies of Pfaff, Klingenberg and Heldt as well as direct evidence obtained by M. Klingenberg and F. Palmieri (unpublished results) with mitochondria devoid of outer membranes (prepared according to Parsons2) show that the exchange between exogenous and endogenous adenine nucleotides^{1,3} occurs at

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